

Trapping of Intermediates in the Interconversion of Heteroyohimbine Alkaloids

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The isolation of (*Z*)-isositsirikines **8** and **9** in the course of NaBH₄ reduction of 19-epicathenamine **2** demonstrates the intermediacy of the (*Z*)-conjugated iminium salt **5** in the interconversion of the heteroyohimbine alkaloids. A 400 MHz ¹H NMR study carried out on compounds **1**, **2**, and **6–9** permitted the determination of all the chemical shifts and most of the coupling constants.

The mechanism of the biosynthetic formation of heteroyohimbine alkaloids is well documented.¹ The intermediacy of the epimeric conjugated iminium salts (*E*)-**3** and (*Z*)-**5** (Scheme 1) has been postulated to explain the formation of cathenamine (19*S*) **1** and epicathenamine (19*R*) **2** heteroyohimbines, respectively.^{2,3}

However, only the *E* alkene intermediate (4,21-dehydrogeissoschizine) **3** has been isolated from *Guettarda eximia* (Rubiaceae).⁴ Moreover, geissoschizine (the 4,21-dihydro derivative of **3**) and the related alkaloids **6** and **7** all have the *E* configuration.

It has been proposed that the less abundant (19*R*)-heteroyohimbine alkaloids are formed by a 1,4-addition of the enol function onto the *Z* alkene derived from the more stable *E* alkene.³ Moreover, the facile formation of epicathenamine **2** from cathenamine **1** in chloroform solution in the presence of alumina* implies the intermediacy of a *Z* alkene during the epimerization at C-19.⁴

* Cathenamine **1** is stable in chloroform solution (in the absence of alumina), whereas epicathenamine **2** slowly epimerises in the same conditions, indicating a lower stability.

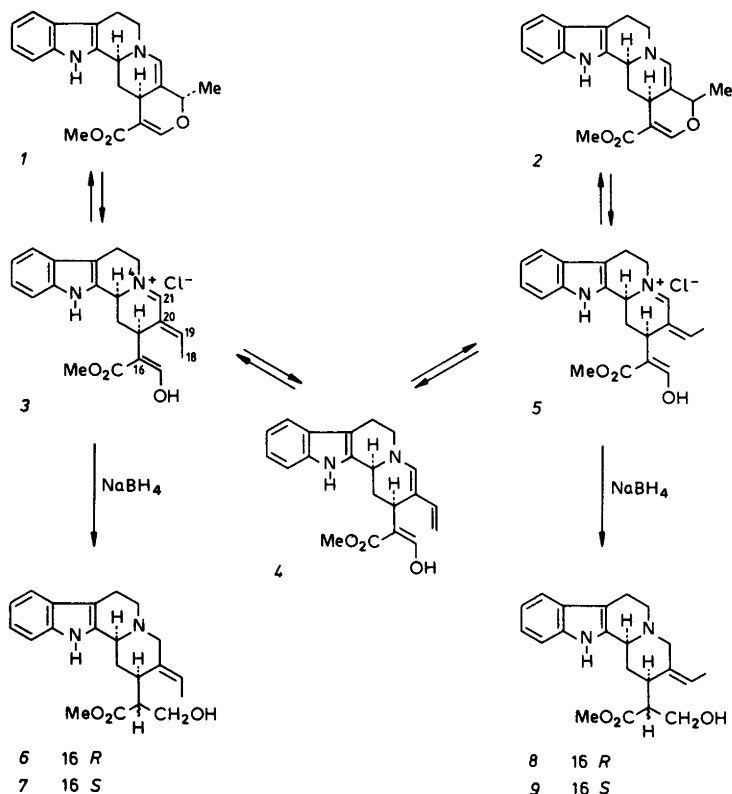
The NaBH₄ reduction of 4,21-dehydrogeissoschizine hydrochloride **3** in methanol has been shown to lead *via* geissoschizine to isositsirikines **6** and **7** (in addition to tetrahydroalstonine), and thus can serve as an indirect method to show the presence of an *E* configurational ethylidene side chain in the initial iminium salt.⁴ It has also been found that the NaBH₄ reductions of cathenamine **1** and epicathenamine **2** yield mainly tetrahydroalstonine and epiajmalicine, respectively.^{3,4}

The NaBH₄ reduction in methanol of the equilibrated mixture obtained from the 4,21-dehydrogeissoschizine hydrochloride **3** (*vide infra*) afforded after purification, besides the expected major products tetrahydroalstonine and 19-epiajmalicine, the known *E* isositsirikines **6** and **7** and more interestingly the *Z* isositsirikines **8** and **9** (Scheme 1).

In these conditions the *E* isositsirikines **6** and **7** cannot be expected to form directly from cathenamine **1**.⁴ Their formation, along with that of the *Z* isositsirikines **8** and **9**, a consequence of trapping the *E* and *Z* iminium salts **3** and **5**, can be interpreted as the results of an equilibrium (Scheme 1).

Although the presence of dienamine **4**, the necessary intermediate between **3** and **5**, could not be experimentally shown (no 18,19-vinyl derivative was isolated after NaBH₄ reduction), its intermediacy has been indirectly shown in a similar epimerization process leading to the yohimbine alkaloids.⁵

To obtain useful ¹H NMR data for the further identification and structure determination of indole alkaloids of similar type we undertook a 400 MHz



Scheme 1.

Table 1. ^1H NMR data of catenamine 1 and epi-catenamine 2. Spectra were run in CDCl_3 at 400 MHz. Values are in δ (TMS=0), s, singlet, d, doublet, t, triplet, q, quartet, m, multiplet, br, broad. The coupling constants between the aromatic protons are not included.

	1	2
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Chemical shifts

H-3	4.30 dd	4.34 dd
H-5 α	3.33 ddd	3.41 ddd
H-5 β	3.28 dd	3.30 dd
H-6 α	2.73 br d	2.73 br d
H-6 β	2.88 ddd	2.88 ddd
H-9	7.46	7.46
H-10	7.08	7.08
H-11	7.16	7.16
H-12	7.32	7.32
H-14 α	3.18 ddd	3.20 ddd
H-14 β	1.46 ddd	1.48 ddd
H-15	3.54 br dd	3.54 br dd
H-17	7.54 d	7.64 d

Table 1. Continued.

H-18	1.42 d	1.48 d
H-19	4.64 br q	4.46 br q
H-21	6.18 dd	6.21 dd
CO ₂ Me	3.72 s	3.70 s
NH	7.98 br s	7.98 br s

Coupling constants, compound 1

$J_{3,14\alpha} = 3$ Hz; $J_{3,14\beta} = 12$ Hz; $J_{5\alpha,5\beta} = 12$ Hz; $J_{5\alpha,6\alpha} \approx 2$ Hz; $J_{5\alpha,6\beta} = 6$ Hz; $J_{5\beta,6\alpha} < 1$ Hz; $J_{5\beta,6\beta} \approx 5$ Hz; $J_{6\alpha,6\beta} = 15$ Hz; $\Pi_{14\alpha,14\beta} = 12$ Hz; $J_{14\alpha,15} = 5$ Hz; $J_{14\beta,15} = 12$ Hz; $J_{15,17} < 0.5$ Hz; $J_{15,21} < 0.5$ Hz; $J_{18,19} = 6.5$ Hz; $J_{19,21} < 0.3$ Hz.

Coupling constants, compound 2

$J_{3,14\alpha} \approx 3$ Hz; $J_{3,14\beta} = 12$ Hz; $J_{5\alpha,5\beta} = 12$ Hz; $J_{5\alpha,6\alpha} \approx 2$ Hz; $J_{5\alpha,6\beta} \approx 6$ Hz; $J_{5\beta,6\alpha} < 1$ Hz; $J_{5\beta,6\beta} \approx 5$ Hz; $J_{6\alpha,6\beta} = 15$ Hz; $J_{14\alpha,14\beta} = 12$ Hz; $J_{14\alpha,15} = 5$ Hz; $J_{14\beta,15} = 12$ Hz; $J_{15,17} < 0.5$ Hz; $J_{15,21} < 0.5$ Hz; $J_{18,19} = 6.5$ Hz; $J_{19,21} < 0.5$ Hz.

Table 2. ^1H NMR data of isositsirikines 6–9. Spectra were run in CDCl_3 at 400 MHz. Values are in δ (TMS=0), s, singlet, d, doublet, t, triplet, q, quartet, m, multiplet, br, broad. The signals due to the OH-groups and the coupling constants between the aromatic protons are omitted.

	6	7	8	9
Chemical shifts				
H-3	4.31 br s	3.9 br s	3.64 br d	3.44 br d
H-5 α	3.15 ddd	2.84 ddd	2.74 m	2.63 ddd
H-5 β	3.27 dd	3.17 dd	3.19 dd	3.11 dd
H-6 α	2.65 br d	2.68 br d	2.76 br d	2.73 br d
H-6 β	3.0 m	3.0 m	3.0 m	3.0 m
H-9	7.48	7.48	7.48	7.45
H-10	7.10	7.09	7.10	7.06
H-11	7.17	7.14	7.16	7.11
H-12	7.38	7.31	7.33	7.28
H-14 α	2.26 ^a m	2.27 ^b m	2.17 ddd	2.26 ddd
H-14 β	2.22 ^a m	2.25 ^b m	1.72 ddd	1.42 ddd
H-15	3.10 m	3.38 m	2.66 m	2.82 m
H-16	2.52 m	2.66 m	3.01 m	2.94 m
H-17	3.55 br dd	3.92 br dd	3.94 br dd	3.95 br dd
H-17'	3.50 br dd	3.87 br dd	3.85 br dd	3.88 br dd
H-18	1.67 d	1.63 d	1.74 d	1.67 d
H-19	5.64 br q	5.52 br q	5.48 br q	5.36 br q
H-21 α	3.54 br d	3.08 br d	2.91 br d	2.76 br d
H-21 β	2.93 br d	3.80 br d	3.82 br d	3.85 br d
CO ₂ Me	3.67 s	3.57 s	3.76 s	3.72 s
NH	8.67 br s	8.23 br s	7.91 br s	8.21 br s

Coupling constants, compound 6

$J_{5\alpha,5\beta}=12$ Hz; $J_{5\alpha,6\alpha}=4$ Hz; $J_{5\alpha,6\beta}=12$ Hz; $J_{5\beta,6\alpha}\approx 1$ Hz; $J_{5\beta,6\beta}=5.5$ Hz; $J_{6\alpha,6\beta}=15$ Hz; $J_{16,17}=7$ Hz; $J_{16,17'}=5$ Hz; $J_{17,17'}=12$ Hz; $J_{18,19}=6.5$ Hz; $J_{19,21\alpha}<0.5$ Hz; $J_{19,21\beta}<0.5$ Hz; $J_{21\alpha,21\beta}=12$ Hz.

Coupling constants, compound 7

$J_{5\alpha,5\beta}=12$ Hz; $J_{5\alpha,6\alpha}=4$ Hz; $J_{5\alpha,6\beta}=12$ Hz; $J_{5\beta,6\alpha}\approx 1$ Hz; $J_{5\beta,6\beta}=5.5$ Hz; $J_{6\alpha,6\beta}=15$ Hz; $J_{16,17}=5$ Hz; $J_{16,17'}=5$ Hz; $J_{17,17'}=12$ Hz; $J_{18,19}=6.5$ Hz; $J_{19,21\alpha}<0.5$ Hz; $J_{19,21\beta}<0.5$ Hz; $J_{21\alpha,21\beta}=12$ Hz.

Coupling constants, compound 8

$J_{3,14\alpha}=4$ Hz; $J_{3,14\beta}=12$ Hz; $J_{5\alpha,5\beta}=12$ Hz; $J_{5\alpha,6\beta}=12$ Hz; $J_{5\beta,6\alpha}\approx 1$ Hz; $J_{5\beta,6\beta}=5.5$ Hz; $J_{6\alpha,6\beta}=15$ Hz; $J_{14\alpha,14\beta}=12$ Hz; $J_{14\alpha,15}=4$ Hz; $J_{14\beta,15}=12$ Hz; $J_{16,17}=7$ Hz; $J_{16,17'}=5$ Hz; $J_{17,17'}=12$ Hz; $J_{18,19}=6.5$ Hz; $J_{19,21\alpha}<0.5$ Hz; $J_{19,21\beta}<0.5$ Hz; $J_{21\alpha,21\beta}=12$ Hz.

Coupling constants, compound 9

$J_{3,14\alpha}=4$ Hz; $J_{3,14\beta}=12$ Hz; $J_{5\alpha,5\beta}=12$ Hz; $J_{5\alpha,6\beta}=12$ Hz; $J_{5\beta,6\alpha}\approx 1$ Hz; $J_{5\beta,6\beta}=5.5$ Hz; $J_{6\alpha,6\beta}=15$ Hz; $J_{14\alpha,14\beta}=12$ Hz; $J_{14\alpha,15}=4$ Hz; $J_{14\beta,15}=12$ Hz; $J_{16,17}=7$ Hz; $J_{16,17'}=5$ Hz; $J_{17,17'}=12$ Hz; $J_{18,19}=6.5$ Hz; $J_{19,21\alpha}<0.5$ Hz; $J_{19,21\beta}<0.5$ Hz; $J_{21\alpha,21\beta}=12$ Hz.

^{a,b} Assignments may be interchanged.

^1H NMR study of the cathenamines 1 and 2, and of the four isositsirikines 6–9.

The application of consecutive double resonance operations permitted all the protons in compounds

1, 2 and 6–9 to be discovered and the coupling constants presented in Tables 1 and 2 to be determined.

The H-3 signal of *E* isositsirikine 6 is a broad

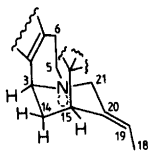


Fig. 1. The predominant conformation of the *E* isositsirikine 6.

singlet (in reality a badly resolved triplet) at δ 4.31 (*cis*-quinolizidine). This seems to indicate that, owing to a strong interaction in the "normal" conformation (*trans*-quinolizidine) between the methyl group of the *E* ethylidene chain and the side chain at C-15, the predominant conformation is the following (Fig. 1):

The 1,3-interaction between the indole nucleus and the C-15 side-chain seems to be less hampering than in geissoschizine.⁶

The same phenomenon, although less pronounced, is present in the 16-*epi E* isositsirikine 7. As would be expected this phenomenon is absent from the *Z* isositsirikines 8 and 9.

The distinction between the *E* isositsirikine 6 and the 16-*epi E* isositsirikine 7 has been made earlier by chemical correlation⁷ and recently by a ¹H NMR study,⁸ in which clearly different chemical shifts were observed for the respective -CH₂- and CH₃COO- groups attached to C-16 (Table 2).

The differences in the corresponding chemical shifts of the isomeric *Z* isositsirikines 8 and 9 (Table 2) are much smaller and do not permit an *a priori* distinction between the two epimers.*

EXPERIMENTAL

The NMR spectra were recorded on a laboratory-constructed 400 MHz ¹H high resolution spectrometer (I.E.F. 400)⁹ and obtained by collecting 8 to 64 free-induction decay signals for a \approx 0.01 M solution of the sample in 450 μ l of CDCl₃. In the case of epicathenamine 2 a 1:1 mixture of cathenamine 1 and epicathenamine 2 was used (*vide infra*).**

One gram of 4,21-dehydrogeissoschizine hydrochloride 3 was equilibrated in 300 ml of chloroform

in the presence of 100 g of alkaline alumina [Merck, aluminium oxide 90 active basic (act.1)] during 24 h. The equilibration leads to a 1:1 mixture of cathenamine 1 and epicathenamine 2 (¹H NMR). The mixture was filtered, the organic solvent evaporated and the obtained residue immediately reduced with NaBH₄ in MeOH containing a trace of acetic acid. After the normal work-up the products (360 mg) were separated by successive TLC (silica gel) treatments yielding besides tetrahydroalstonine and 19-*epiajmalicine*, *E* isositsirikine 6 and 16-*epi E* isositsirikine 7, which were identified by direct comparison (TLC, IR, ¹H NMR, MS) with authentic samples, as well as *Z* isositsirikine 8 and 16-*epi Z* isositsirikine 9 (¹H NMR, MS).

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* The presentation of *Z* isositsirikines in formulas 8 and 9 is arbitrary and could be reversed.

** As we did not succeed in isolating pure epicathenamine 2 we decided to use a 1:1 mixture of cathenamine 1 and epicathenamine 2.